



## Genetic Diversity of Purslane Populations of Iran Based on Some Morphological and Biochemical Traits

**Running title:** Genetic Diversity of Purslane Populations of Iran

**Daryush Talei<sup>1\*</sup>, Amir Mohammad Naji<sup>2</sup> and Narjess Labbaf<sup>2</sup>**

<sup>1</sup>Medicinal Plants Research Center, Shahed University, Tehran, Iran

<sup>2</sup>Department of Biotechnology, Agriculture Faculty, Shahed University, Tehran, Iran

Article History: Received: 12 May 2020/Accepted in revised form: 21 June 2020

© 2012 Iranian Society of Medicinal Plants. All rights reserved.

### Abstract

*Portulaca oleracea* L. is a medicinal plant belonging to Portulacaceae family, which exhibited a wide scope of pharmaceutical properties such as pain killer, antipyretic, anti-viral and anti-fungal. The aim of the present study was evaluation of morphological diversity 18 populations of *P. oleracea*. The seeds of 18 populations were cultivated based on a completely randomized design with three replicates in the research farm of Medicinal Plants Research Center, Shahed University, Tehran, Iran. The morphological traits, chlorophyll and protein contents were measured in the vegetative stage. The analysis of variance showed that there was a significant difference among populations of *P. oleracea* in terms of the most studied morphological and biochemical traits. The highest shoot dry weight ( $84.84 \pm 12.70$  g per plant) belonged to the Zanjan population, while the lowest shoot dry weight ( $18.89 \pm 1.72$  g per plant) belonged to the population of Lourdgan. The correlation between most of the studied traits was positively significant. The cluster analysis of the 18 *P. oleracea* populations based on morphological and biochemical traits produced three main clusters. Overall, the outcomes of the present study were indicated the presence of high genetic variability among the *P. oleracea* populations. Our findings suggested that collection of the plants from different regions can be used for hybridization to generate useful recombinants in the segregating generations and improve breeding varieties of *P. oleracea* and can be utilized for preservation and maintenance the germplasm of this medicinal plant.

**Keywords:** Biochemical, Genetic diversity, Morphological, *Portulaca oleracea*.

### Introduction

*Portulaca oleracea* commonly known as Purslane is an herbaceous annual in the family of Portulacaceae. The plant is distributed all over the world including Europe, America, Canada, India, New Zealand, Australia, China and Japan [1]. It is particularly well adapted to the warm, moist conditions found in irrigated agricultural and ornamental sites. The plant requires a moist light rich well-drained soil in a sunny position. The plants will not produce good quality leaves when growing in dry conditions. The plants take about six to eight weeks to produce a crop from seed and

can then be harvested on a cut and come again principle, providing edible leaves for most of the summer. The plant has a round, smooth, procumbent, succulent stem, with small, oblong, wedge-shaped, sub-sessile, alternate or sub-opposite dark-green leaves. The flowers are small, yellow, solitary or clustered, stalkless, placed above the last leaves on the branches, blooming in June and July, and opening only for a short time towards noon. Seeds are reddish brown to black, oval, and tiny [2,3].

The plant is rich in fatty acids, proteins, and vitamins (mainly vitamin A, C, E, and some vitamin B and carotenoids), as well as dietary

\*Corresponding author: Medicinal Plants Research Center, Shahed University, Tehran, Iran  
Email Address: d.talei@shahed.ac.ir

minerals, such as magnesium, calcium, potassium, and iron with about 70% of its fatty acids being unsaturated and about 50% containing only omega-3 fatty acids. At the present there are two types of betalain alkaloid pigments, the reddish betacyanins and the yellow betaxanthins [4]. According to Iranian traditional medicine sources, purslane is an antinociceptive, anesthetic, antiseptic, anti-ascorbate, anti-inflammatory, anti-inflammatory, blood purifier, anti-fungal and reducing swelling and abscesses, insect bite and scorpion sting [5].

Evaluation of genetic variation of plants is one of the important requirements for plants in order to achieve high yield and desirable quality. In breeding programs, genetic diversity is the first step that has been created over the years and is one of the most important factors in the survival of creatures, including plants, for adaptation to environmental changes [6]. Genetic diversity can be estimated based on genealogy analyzes using various methods including morphological traits, protein markers, and molecular markers [7,8]. Initially, morphological and agronomic characteristics are often used for basic characterization [9]. Interspecific and intergeneric hybridisation has been the major source of genetic variation in breeding new cultivars of ornamental plants [10,11]. With respect to genetic diversity, several proteins and molecular polymorphic have been reported in the purslane (*Portulaca oleracea* L.) accessions [12-14].

*P. oleracea* is a very variable species, cytologically, *P. oleracea* is characterized by a sequence of polyploids, from a base number of  $x=9$ . The variability expressed by the existence of diploid ( $2n=18$ ), tetraploid ( $2n=36$ ) and hexaploid ( $2n=54$ ) populations. Based on seed size and seed-coat cell morphology, up to ten subspecies has been differentiated. The cultivated forms ( $2n=54$ ), which usually have taller plants and larger seeds, can best be distinguished botanically at cultivar level [15, 16].

In *P. oleracea*, genetic diversity of 10 purslane accessions has been evaluated using AFLP markers [17]. Alam *et al.* [18] evaluated 45 Malaysian accessions of *P. oleracea* using EST-SSR marker and reported 71.87% variation within population and 28.13% among populations. Also, Alam *et al.* [18] in a study on genetic diversity of 45 *P. oleracea* accessions using ISSR marker reported 89% variation within population and 11% among populations. Each marker explores a part of the

differences and similarities within the genome. Because of the commercial and pharmaceutical importance of this plant and the growing trend of its use and cultivation, improving the plant with high yield and secondary metabolites cultivars using wild plants or selected ecotypes as well as conservation of natural germplasm resources in the country seems necessary. The objective of this study was to evaluate morphological and biochemical diversity among 18 populations of *P. oleracea* collected from different states of Iran to estimate genetic relationships among the population which can be utilized for breeding program and phytochemical study.

## Material and Methods

### Plant Materials

Seeds of 18 populations of *P. oleracea* seeds were collected from different regions of Iran (Table 1).

### Growth Conditions

The seeds were surface sterilized with 10% sodium hypochlorite (NaClO) solution for 10 minutes and thoroughly rinsed with distilled water [19]. The seeds of 18 populations were cultivated based on completely randomized design with three replicates in the Research farm of Medicinal Plants Research Center, Shahed University, Tehran, Iran. Each population was cultivated in  $1 \times 1.2$  m plots with three rows and 25 cm spacing and a density of 20-30 plants per plot. The plants were irrigated every day until flowering stage. Before flowering stage ten plants were randomly selected from each experimental unit and the data on morphological and biochemical traits such as average shoot length (cm), stem diameter (mm), number of branches, stem color, shoot fresh weight (g per plant), shoot dry weight (g per plant) and chlorophyll ( $\mu\text{g/g}$  FW) and protein contents ( $\mu\text{g/g}$  FW) were measured. The dry shoot weights after drying at the 68 °C for 48 hours were measured.

### Determination of Chlorophyll and Protein Content

The chlorophyll content of leaves was measured using Chlorophyll-Meter-XT-SPAD-502 equipment (Konica Minolta, Inc., Tokyo, Japan) [20]. To determine the protein content, one gram fresh leaf from the 18 different *P. oleracea* population was grounded in liquid nitrogen using pre-cooled mortar and pestle to obtain a fine powder and then homogenized with 2 mL of the

HEPES/KOH buffers according to the method of Talei *et al.*, [21]. Finally, the total protein concentration was determined by the Bradford method [22] at 595 nm, using a spectrophotometer (Lambda 25, UV/VIS).

#### Statistical Analysis

The analysis of variance and Duncan's multiple range tests were performed using SPSS25 software

at significance level of P 0.01, and the cluster analysis and PCA analysis were performed using JMP software version 13. The formula used to estimate the broad-sense heritability is [23]:

$$h^2 = \frac{\delta_G^2}{\delta_P^2} \text{ and } \delta_P^2 = \delta_G^2 + \delta_e^2$$

Where  $\delta_G^2$  is the genetic variance,  $\delta_P^2$  is the phenotypic variance and  $\delta_e^2$  is the environmental variance.

**Table 1** Geographical origins of the of *Portulaca oleracea* L. population from different part of Iran.

code	Region originated	Latitude	Longitude	Altitude (m)
1	Varamin	35° 12' 36.513"	51° 40' 26.832"	842
2	Malard	35° 41' 29.949"	50° 50' 2.720"	1159
3	Karaj	35° 49' 11.727"	50° 56' 11.296"	1282
4	Qom	34° 37' 35.903"	50° 55' 37.719"	911
5	Abhar	36° 10' 29.042"	49° 15' 20.246"	1574
6	Zanjan	36° 38' 41.781"	48° 30' 54.880"	1642
7	Qazvin	36° 17' 52.241"	50° 2' 41.569"	1338
8	Bandar Turkman	36° 49' 45.880"	54° 3' 19.337"	-26
9	Talesh	37° 52' 26.749"	48° 54' 24.150"	-6
10	Hamedan	34° 49' 52.081"	48° 30' 56.491"	1756
11	Razan	35° 23' 18.355"	49° 1' 29.424"	1839
12	Izeh	29° 28' 0.801"	51° 16' 18.382"	102
13	Babol	36° 29' 39.799"	52° 42' 25.769"	8
14	Tonekabon	36° 46' 15.806"	50° 50' 49.824"	48
15	Lourdgan	30° 39' 18.529"	51° 36' 53.288"	1843
16	Boroujerd	33° 52' 32.840"	48° 45' 32.557"	1533
17	Kashan	33° 57' 12.852"	51° 22' 9.576"	1015
18	Folad Shahr	32° 27' 55.425"	51° 24' 13.244"	1708

**Table 2** Variance analysis of 18 purslane populations in terms of some morphological and biochemical traits.

S.O.V	df	Plant height	Stem diameter	Branches number	Fresh weight	Dry weight	Chlorophyll content	Protein content
Rep	2	727.04 **	5.53 *	4.22 **	2301.30 <sup>ns</sup>	620.78 <sup>ns</sup>	195.339 **	0.022 <sup>ns</sup>
Genotype	17	62.06 **	3.01 **	0.41 <sup>ns</sup>	31588.23 **	822.50 **	40.705 <sup>ns</sup>	0.016 *
Error	34	36.82	2.06	0.26	7484.56	116.58	35.024	0.007
CV (%)		9.62	15.99	15.93	25.83	23.62	12.80	18.93

\*, \*\*: Significant at 5% and 1% probability level, and ns; non-significant.

## Result and Discussion

The results of analysis of variance showed that there is a significant difference between the purslane populations based on most of the studied

traits (P 0.01), while there was no significant difference between the population based on number of branches and chlorophyll content (Table 2).

The highest ( $70.57 \pm 3.12$  cm) and lowest plant height ( $54.85 \pm 2.35$  cm) belonged to Abhar population from the Zanjan state and Bandar Turkman population from the Golestan state, respectively. The highest stem diameter ( $10.91 \pm 0.32$  mm) belonged to the Boroujerd population from the Lorestan state, while the lowest stem diameter ( $7.55 \pm 0.30$  mm) belonged to the population of Bandar Turkman.

The results indicated that the highest ( $84.84 \pm 12.70$  g per plant) and the lowest shoot dry weight ( $18.89 \pm 1.72$  g per plant) belonged to the Zanjan and Lourdgan populations, respectively. The results also showed that the highest protein content ( $0.583 \pm 0.04$   $\mu\text{g}\cdot\text{g}^{-1}$  FW) belonged to the Bandar Turkman population, while the lowest protein content ( $0.306 \pm 0.04$   $\mu\text{g}\cdot\text{g}^{-1}$  FW) belonged to the Zanjan population. There were no significant differences among different population in terms of chlorophyll contents, nevertheless chlorophyll content variation due to different population was

between 40.98 to  $55.80$   $\mu\text{g}\cdot\text{g}^{-1}$  FW (Table 3). Therefore, according to the results, it can be said that there are many differences in measured traits among the populations, which can be used for breeding programs and selection of the desired genotypes.

Mohebodini, *et al.*, [13] studied twenty purslane plant populations for morphological and agronomic traits. The reason for the high variability among the purslane in terms of measured traits could be the pursuit of free purslane pollination and the possibility of pollen transfer between the species [24]. The correlation between the most morphological traits was very significant and positive, while the correlation between protein content and plant height, and fresh and dry shoot weight was significantly negative. The highest correlation was found between shoot fresh weight and shoot dry weight ( $r = 0.94$ ), and the lowest correlation between number of branches and protein ( $r = 0.25$ ) (Table 4).

**Table 3** Mean comparison of different purslane populations based on some studied traits.

Genotype	Plant height (cm)	Stem diameter (mm)	Fresh weight (g)	Dry weight (g)	Protein content ( $\mu\text{g}/\text{g}$ FW)
Varamin	$64.50 \pm 2.15$ a-d	$10.44 \pm 0.30$ ab	$424.37 \pm 30.21$ ab	$71.46 \pm 3.76$ ab	$0.39 \pm 0.07$ bc
Malard	$64.00 \pm 4.57$ a-d	$8.51 \pm 0.31$ a-c	$187.33 \pm 53.18$ de	$48.46 \pm 7.86$ c-e	$0.44 \pm 0.08$ a-c
Karaj	$67.13 \pm 4.97$ a-d	$9.52 \pm 0.28$ a-c	$479.60 \pm 21.66$ a	$45.45 \pm 3.88$ c-e	$0.40 \pm 0.07$ bc
Qom	$64.03 \pm 5.73$ a-c	$8.66 \pm 0.18$ a-c	$246.33 \pm 61.74$ c-e	$35.34 \pm 7.82$ d-f	$0.39 \pm 0.09$ bc
Abhar	$70.57 \pm 3.12$ a	$8.89 \pm 0.45$ a-c	$255.93 \pm 19.75$ c-e	$46.98 \pm 3.95$ c-e	$0.40 \pm 0.08$ bc
Zanjan	$67.41 \pm 3.61$ a-c	$7.85 \pm 0.31$ bc	$357.30 \pm 51.92$ a-c	$84.84 \pm 12.70$ a	$0.31 \pm 0.04$ c
Qazvin	$64.67 \pm 0.99$ a-d	$9.07 \pm 0.37$ a-c	$187.50 \pm 7.47$ de	$47.53 \pm 3.07$ c-e	$0.49 \pm 0.01$ ab
Bandar Turkman	$54.85 \pm 2.35$ d	$7.55 \pm 1.09$ c	$233.03 \pm 50.17$ c-e	$55.08 \pm 7.81$ b-d	$0.58 \pm 0.04$ a
Talesh	$59.48 \pm 1.49$ a-d	$8.94 \pm 0.67$ a-c	$190.47 \pm 30.15$ c-e	$60.54 \pm 6.11$ bc	$0.52 \pm 0.03$ ab
Hamedan	$68.20 \pm 4.77$ ab	$8.75 \pm 0.33$ a-c	$229.60 \pm 2.66$ c-e	$29.46 \pm 0.59$ e-f	$0.47 \pm 0.05$ ab
Razan	$58.07 \pm 6.21$ b-d	$9.81 \pm 0.72$ a-c	$194.27 \pm 29.13$ c-e	$23.83 \pm 5.51$ f	$0.53 \pm 0.06$ ab
Izeh	$59.10 \pm 1.81$ a-d	$7.77 \pm 0.44$ bc	$173.13 \pm 28.12$ de	$24.86 \pm 4.32$ f	$0.48 \pm 0.05$ ab
Babol	$68.57 \pm 5.98$ ab	$7.86 \pm 0.27$ bc	$234.73 \pm 24.61$ c-e	$54.68 \pm 2.69$ b-d	$0.36 \pm 0.02$ bc
Tonekabon	$55.87 \pm 3.85$ cd	$10.70 \pm 1.84$ a	$155.10 \pm 30.41$ de	$44.96 \pm 4.47$ c-e	$0.41 \pm 0.02$ bc
Lourdgan	$58.80 \pm 2.08$ a-d	$8.19 \pm 0.65$ a-c	$135.87 \pm 15.09$ e	$18.89 \pm 1.72$ f	$0.37 \pm 0.02$ bc
Boroujerd	$61.63 \pm 3.84$ a-d	$10.91 \pm 0.78$ a	$441.83 \pm 68.98$ ab	$47.26 \pm 8.34$ c-e	$0.47 \pm 0.03$ ab
Kashan	$64.90 \pm 1.24$ a-d	$9.27 \pm 0.86$ a-c	$302.57 \pm 71.80$ b-e	$45.05 \pm 8.91$ c-e	$0.52 \pm 0.02$ ab
Folad Shahr	$63.73 \pm 5.00$ a-d	$8.70 \pm 0.80$ a-c	$319.43 \pm 66.58$ b-d	$38.07 \pm 7.36$ d-f	$0.47 \pm 0.03$ ab

Mean values  $\pm$  SE are from three independent replicates and values superscripted by different letters are significantly different by Tukey's multiple range tests ( $P < 0.01$ ).

**Table 4** Correlations among some morphological and biochemical traits in purslane.

Traits	Plant height	Stem diameter	Branches number	Shoot fresh weight	Shoot dry weight	Chlorophyll
Stem diameter	-0.34	-	-	-	-	-
Branches No	-0.36	0.64**	-	-	-	-
Shoot fresh weight	0.70**	0.66**	0.73**	-	-	-
Shoot dry weight	0.58**	0.38	0.47	0.94**	-	-
Chlorophyll	0.51*	0.31	-0.51*	0.43	-0.42	-
Protein	-0.63**	0.38	-0.25	-0.52*	-0.51*	-0.47

\*\* , \* = Significant at 1 and 5% probability levels.

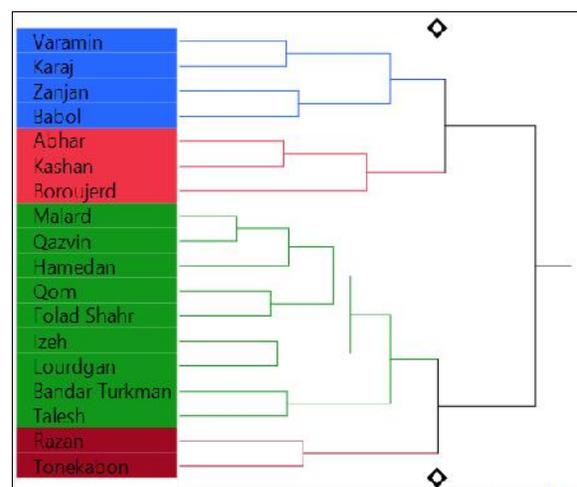
Mohebodini, *et al.*, [13] in a study of 20 different genotypes of purslane, reported that there was a significant positive correlation between plant height, tiller number, leaf length and width, internode length and essential oil, but negative correlation with leaf width. Alam, *et al.*, [12] in a study of 45 different populations collected from different states of Malaysia showed that there was a high level of genetic diversity in the germplasm.

#### Cluster Analysis

Cluster analysis was performed to classify the genotypes into homogenous groups based on the similarity/dissimilarity percentage between them. The unweighted pair group method with arithmetic mean (UPGMA) cluster analysis of the 18 populations of *P. oleracea* based on morphological and biochemical traits produced four clusters. Cluster 1 contained four populations (Varamin, Karaj, Zanjan and Babol), cluster 2 contained three populations (Abhar, Kashan and Boroujerd), cluster 3 contained nine populations (Malard, Qazvin, Hamedan, Qom, Folad Shahr, Izeh, Lourdgan, Bandar Turkman and Talesh) and cluster 4 including two populations (Razan and Tonekabon) (Figure 2). The distance between leader and joiner accessions in cluster analysis based on all measured traits was 5.29. The highest distance (5.29) was obtained among Malard and Varamin accessions, while the lowest distance (0.86) was obtained among Malard and Qazvin accessions. Clustering pattern confirmed that the genetic diversity is independent of a geographical area as accessions collected from different regions were accumulated in same clusters and those from the same region were also distributed into different clusters randomly. The findings of research suggest that plants belonging to different clusters can be used in the hybridization program to produce recombinant

genotyping, genetic diversity, and reproductive breeding programs for *P. oleracea* population.

Alam *et al.*, [25] studied the morphological physiological and mineral traits of 45 Malaysian *P. oleracea* populations and finally divided their cluster analysis into five general groups. The pattern of population variation in this study partly follows the geographical pattern, and the external masses are clustered in the central and western provinces. In another study, the diversity of 9 Iranian purslane populations with 2 foreign populations from Germany and Japan was investigated for morphological and phytochemical characteristics. Cluster decomposition results using WARD algorithm grouped them into four different groups. Qazvin population had more genetic diversity than other populations.



**Fig. 2** A dendrogram generated by UPGMA clustering method of 18 populations of *Portulaca oleracea* L. based on all measured traits.

#### Principal Component Analysis (PCA)

Principal Component Analysis (PCA) is one of the multivariate analyses which reduces the more

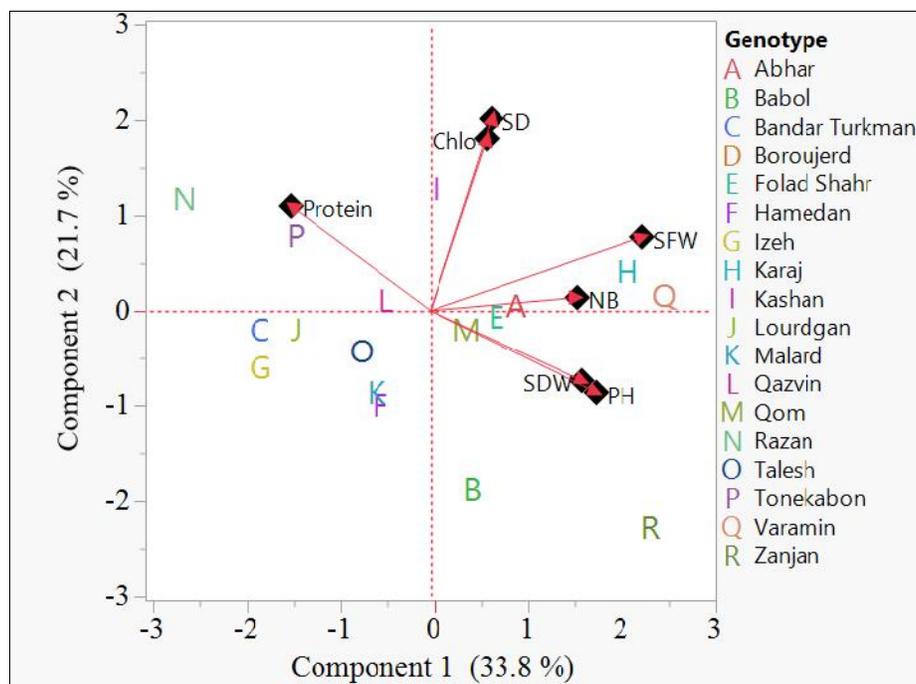
number of correlated variables to limited number uncorrelated individual variables known as Principal Components (PC), thereby, simplifies the multivariate data set. Therefore, for a better understanding of the effectiveness of the studied traits, principal components analysis (PCA) was performed using seven characteristics on the biplot (Figure 3). The PCA revealed two principal components (PC) with Eigen value ranging from 0.21 to 2.36, which made up 55.5% of the total data variance (Figure 3). The PCA also indicated that Babol and Zanjan populations had more plant height and shoot dry weight, while Razan and Tonekabon populations had more protein content. In addition, the PCA results confirmed the results of the clustering. The PCA revealed that the most important character in PC1 was shoot fresh weight and plant height, while chlorophyll content was the least effective, while the most important character in PC2 was stem diameter and chlorophyll content, while number of branches was the least effective (Table 5). Nevertheless, since PC1 and PC2 represented 33.8% and 21.7% of the total variance respectively, so, the importance of the mentioned traits in PC1 and PC2 should be further looked into in future studies.

**Table 5** Component matrix of morphological and biochemical traits in 18 populations of *Portulaca oleracea* L.

Variables	PCA1	PCA2	PCA3
Shoot fresh weight	0.861	0.295	-
Plant height	0.674	-0.33	-
Shoot dry weight	0.616	-0.282	-
Number of branches	0.598	0.053	-
Protein	-0.567	0.419	-
Stem diameter	0.248	0.768	-
Chlorophyll	0.229	0.688	-
Eigen value	2.358	1.508	0.944
Variance%	33.79	21.74	13.49
Cumulative Variance%	33.79	55.53	69.09

Broad-sense Heritability

Broad-sense heritability calculated for all characters revealed that among morphological and biochemical characters, the broad-sense heritability ( $h^2$ ) of shoot dry weight was the highest (66.87) and chlorophyll content was the lowest (5.15) (Table 4). Bhatti *et al.* [26] have reported the higher heritability of shoot and root length in *Gossypium hirsutum* L.



**Fig. 3** The principal components analysis (PCA) of studied traits in eighteen population of *Portulaca oleracea* L. The PCA revealed two principal components (PC). Plant height (PH), stem diameter (SD), number of branches (NB), shoot fresh weight (SFW), shoot dry weight (SDW), chlorophyll (Chlo).

**Table 4** Components of variance and broad-sense heritability of morphological and biochemical characters in eighteen population of *Portulaca oleracea* L.

S.O.V	Plant height	Stem diameter	Branches number	Fresh weight	Dry weight	Chlorophyll content	Protein content	E(MS)
Genotype	62.06**	3.01**	0.41 <sup>ns</sup>	31588.23**	822.50**	40.71 <sup>ns</sup>	0.016*	$\uparrow_e^2 + r\uparrow_g^2$
Error ( $\sigma_e^2$ )	36.82	2.06	0.26	7484.56	116.58	35.02	0.007	$\uparrow_e^2$
$\sigma_p^2$	8.41	0.32	0.05	8034.56	235.31	1.90	0.003	
$\sigma_B^2$	45.23	2.38	0.31	15519.12	351.89	36.92	0.01	
$h_B^2$	18.59	13.45	16.13	51.77	66.87	5.15	30.00	

$\sigma_G^2$ : Genetic variance,  $\sigma_p^2$ : phenotypic variance and  $h_B^2$ : broad-sense heritability.

The present results confirm high heritability of shoot and root dry weight and total dry weight among agronomic traits as well as plant height and number of leave. Therefore, the underlying genetic mechanisms can be considered as direct criterion for assessing genetic diversity.

## Conclusion

In general, the results of the study revealed that there was a high genetic diversity between the *P. oleracea* populations provides important baseline data and a better understanding of conservation, management, and collection strategies for germplasm of this species. The findings of research indicated that the geographically different populations are placed in identical clusters. Also, in general, there is no relation between geographical position and population relatives. Therefore, the findings of research suggest that plants belonging to different clusters can be used in the hybridization program to produce recombinant genotyping, genetic diversity, and reproductive breeding programs for *P. oleracea* populations.

## References

- Movahedian A, Ghannadi A, Vashirnia M. Hypocholesterolemic effects of purslane extract on serum lipids in rabbits fed with high cholesterol levels. *Int J Pharmacol.* 2007;3:285-9.
- Banerjee G, Mukherjee A. Pharmacognostic studies on *Portulaca oleracea* L. leaf. *J Econ Taxon Botany.* 2003;19:69-77.
- Holm LG, Plucknett DL, Pancho JV, Herberger JP. The world's worst weeds. Distribution and biology: University press of Hawaii. 609 pages. 1977.
- Masoodi MH, Ahmad B, Mir SR, Zargar BA, Tabasum N. *Portulaca oleracea* L. a review. *J Pharma Res.* 2011;4:3044-8.
- Zargari A. Medicinal Plants University of Tehran Press. Tehran (in Persian). 1997;3:80-8.
- Mohammadi M, Karimizadeh R, Shefazadeh M, Sadeghzadeh B. Statistical analysis of durum wheat yield under semi-warm dry land condition. *Australian J Crop Sci.* 2011;5:1292-7.
- Chahal G, Gosal S. Principles and procedures of plant breeding: Biotechnological and conventional approaches: Alpha Science Int'l Ltd. 2002.
- Talei D, Valdiani A, Abdullah MP. Impact of protein diversification on morphometric behavior of *Andrographis paniculata* Nees. *Plant Systematics and Evolution.* 2014;300:1003-10. DOI 10.7/s00606-013-0938-z.
- Okpul T, Mace E, Godwin I, Singh D, Wagih M. Evaluation of variability among breeding lines and cultivars of taro (*Colocasia esculenta*) in Papua New Guinea using ISSR fingerprinting and agromorphological characterization. *Plant Genetic Res Newsletter.* 2005;143:8.
- Kaneko Y, Bang SW. Interspecific and intergeneric hybridization and chromosomal engineering of Brassicaceae crops. *Breeding science.* 2014;64:14-22.
- Van Tuyl JM, Lim K-B, editors. Interspecific hybridisation and polyploidisation as tools in ornamental plant breeding. XXI International Eucarpia Symposium on Classical versus Molecular Breeding of Ornamentals-Part I 612. 2003.
- Alam MA, Juraimi AS, Rafii MY, Hamid AA, Arolu IW, Latif MA. Genetic diversity analysis among collected purslane (*Portulaca oleracea* L.) accessions using ISSR markers. *Comptes rendus biologiques.* 2015;338:1-11.
- Mohebodini M, Behnamian M, Dezhsetan S. Assessment of genetic diversity of purslane (*Portulaca oleracea* L.) accessions in Iran: University of Mohaghegh Ardabili. 2016.
- Mosquera SME. Purslane (*Portulaca oleracea* L.) an excellent source of: McGill University. 2013.
- Grubben G. Plant Resources of Tropical Africa (PROTA): Prota. 2008.

16. Matthews JF, Ketron DW, Zane SF. The biology and taxonomy of the *Portulaca oleracea* L.(Portulacaceae) complex in North America. *Rhodora*. 1993;166-83.
17. Ren S, Weeda S, Akande O, Guo Y, Rutto L, Mebrahtu T. Drought tolerance and AFLP-based genetic diversity in purslane (*Portulaca oleracea* L.). *J Biotech Res*. 2011;3:51.
18. Alam MA, Juraimi AS, Rafii M, Hamid AA, Arolu IW, Latif M. Application of EST-SSR marker in detection of genetic variation among purslane (*Portulaca oleracea* L.) accessions. *Brazilian J Botany*. 2015;38:119-29.
19. Talei D, Mihdzar AK, Khanif MY, Saad MS, Valdiani AR. Effects of different surface sterilizers on seed germination and contamination of king of bitters (*Andrographis paniculata* Nees.). *American-Eurasian J Agric Environ Sci*. 2011;10:639-43.
20. Ling Q, Huang W, Jarvis P. Use of a SPAD-502 meter to measure leaf chlorophyll concentration in *Arabidopsis thaliana*. *Photosynthesis Res*. 2011;107:209-14.
21. Talei D, Valdiani A, Puad M. An effective protein extraction method for two-dimensional electrophoresis in the anticancer herb (*Andrographis paniculata* Nees.). *Biotechnology and Applied Biochemistry*. 2013;60:521-6. DOI: 10.1155/2013/319047.
22. Bradford MM. A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding. *Analytical Biochemistry*. 1976;72:248-54.
23. Xu NW, Xu S, Ehlers J. Estimating the broad-sense heritability of early growth of cowpea. *Inter Journal of Plant Genomics*. 2009;2009.
24. Wickramasinghe P, Harrison DK, Johnston ME. Reproductive biology and intergeneric breeding compatibility of ornamental *Portulaca* and *Calandrinia* (Portulacaceae). *Australian J botany*. 2010;57:697-707.
25. Alam MA, Juraimi AS, Rafii M, Hamid AA, Uddin MK, Alam M, et al. Genetic improvement of Purslane (*Portulaca oleracea* L.) and its future prospects. *Mol Biol Reports*. 2014;41:7395-411.
26. Bhatti MA, Azhar F. Salt tolerance of nine *Gossypium hirsutum* L. varieties to NaCl salinity at early stage of plant development. *Int J Agric Biol*. 2002;4:544-6.